

# Fungal Contamination in Schools

## Key Insights and Assessment Strategies

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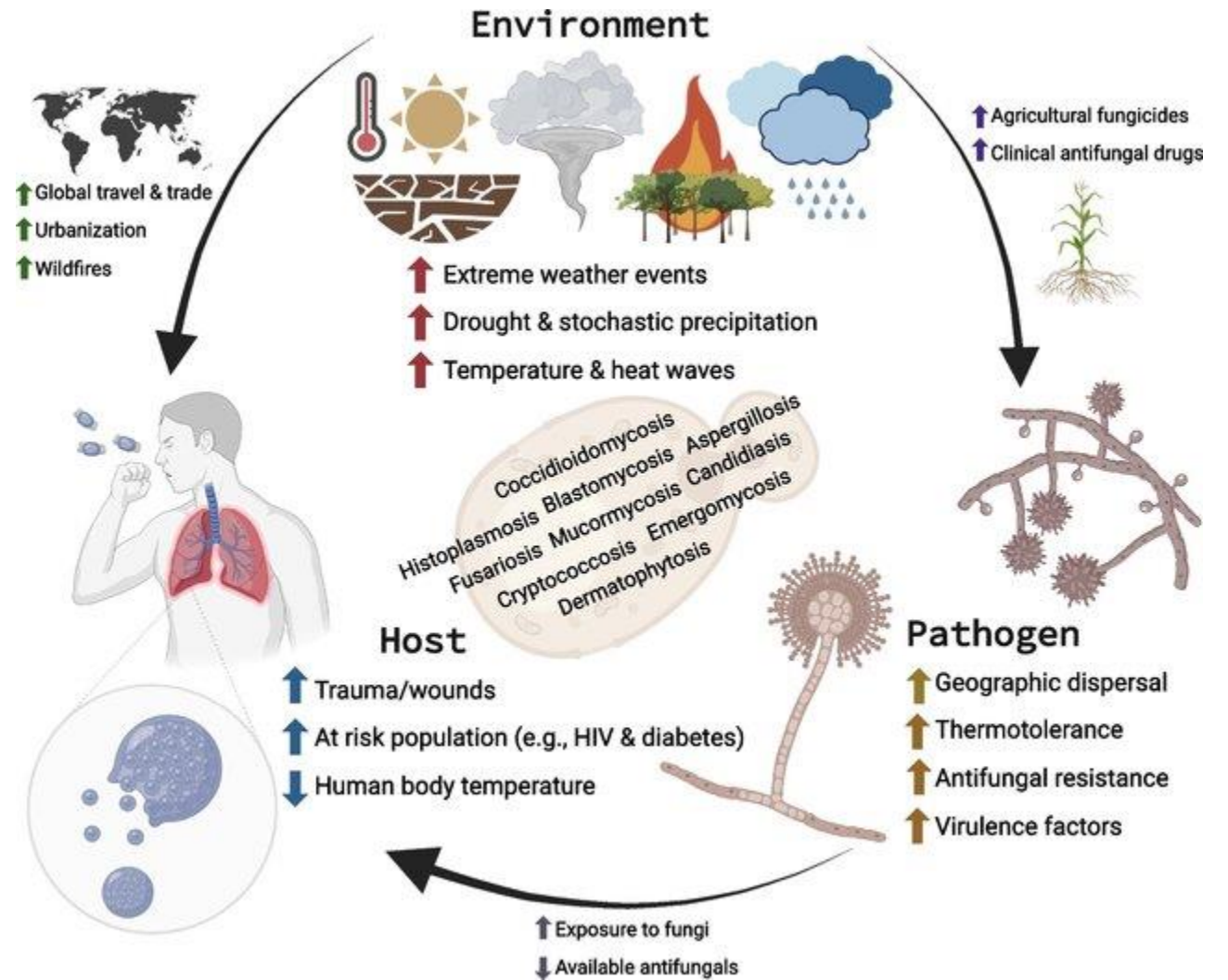
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# Introdução

**Global Health Threat:** Antimicrobial resistance and fungal pathogens (WHO, 2022).

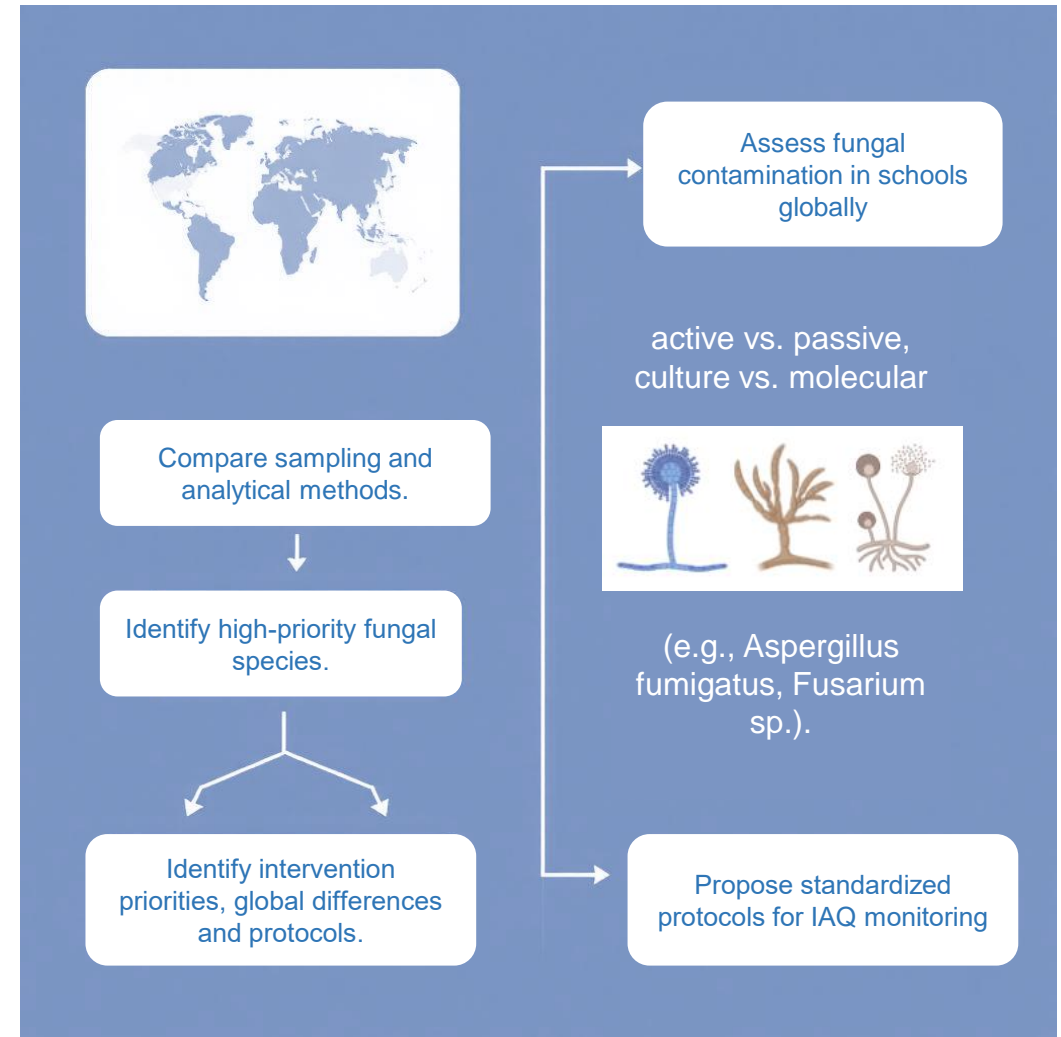
**Climate Change Impact:** Warmer, wetter climates increase fungal dispersal and resistance (Seidel et al., 2024).

- **Focus:** Schools as high-risk environments for fungal exposure and health effects (respiratory issues, allergies, cognitive impacts)

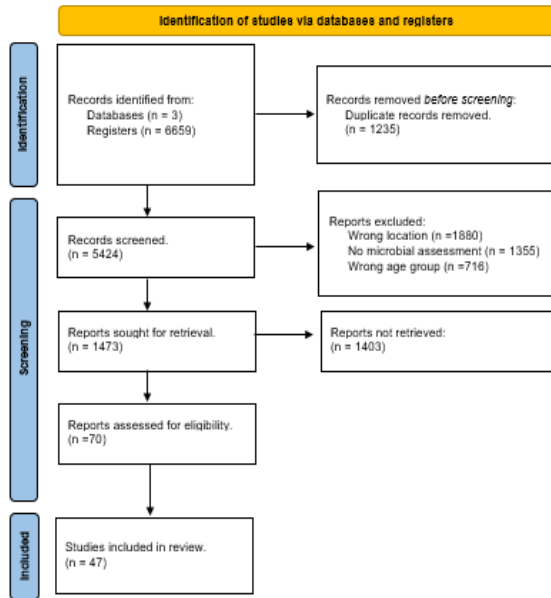


# Objectives

- Assess fungal contamination in schools globally.
- Compare sampling and analytical methods (active vs. passive, culture vs. molecular).
- Identify high-priority fungal species (e.g., *Aspergillus fumigatus*, *Fusarium* sp.).
- Propose standardized protocols for IAQ monitoring.



# Methodology



## Systematic review



**Data Sources**  
PubMed, Scopus,  
Web of Science  
(2010–2024).

**Inclusion Criteria**  
English, original  
studies, elementary  
schools, fungal focus.

**6,659** articles  
screened  
→ **47 selected**  
for analysis.

**Variables Extracted:**  
Sampling methods,  
fungal species, climatic  
conditions, health  
outcomes.

# Geographic Distribution

## Europe

Portugal (N=3)

Finland (N=8)

Spain (N=2)

Netherlands (N=2)

Denmark (N=4)

Sweden (N=2)

Greece (N=1)

Italy (N=1)

Norway (N=1)

France (N=1)

Turkey (N=3)

England (N=1)

Germany (N=1)

Poland (N=1)

Lithuania (N=1)

## North America

USA (N=12)

Canada (N=2)

## Asia

China (N=2)

Taiwan (N=3)

South Korea (N=2)

Saudi Arabia (N=1)

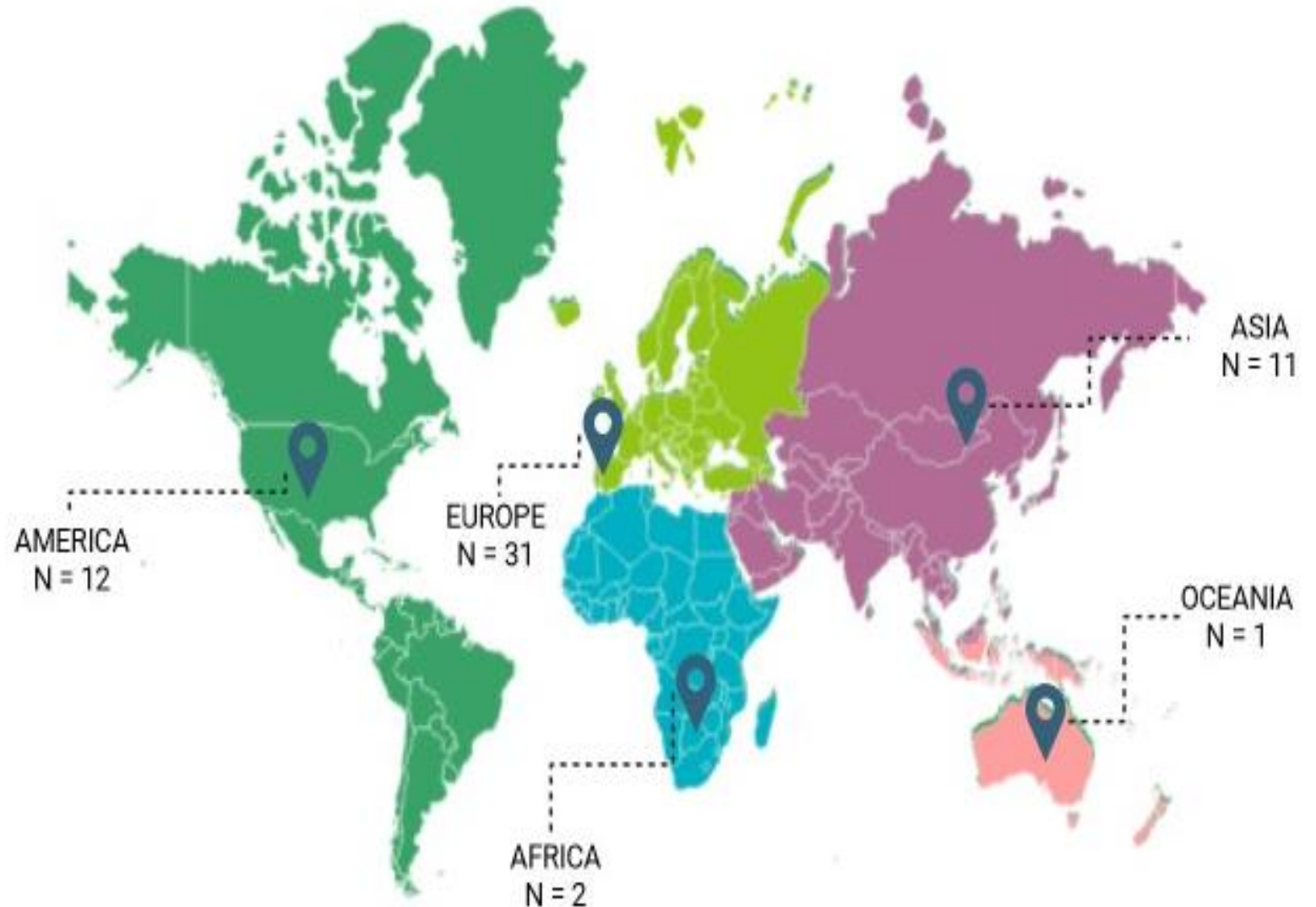
## Africa

Egypt (N=1)

Ethiopia (N=1)

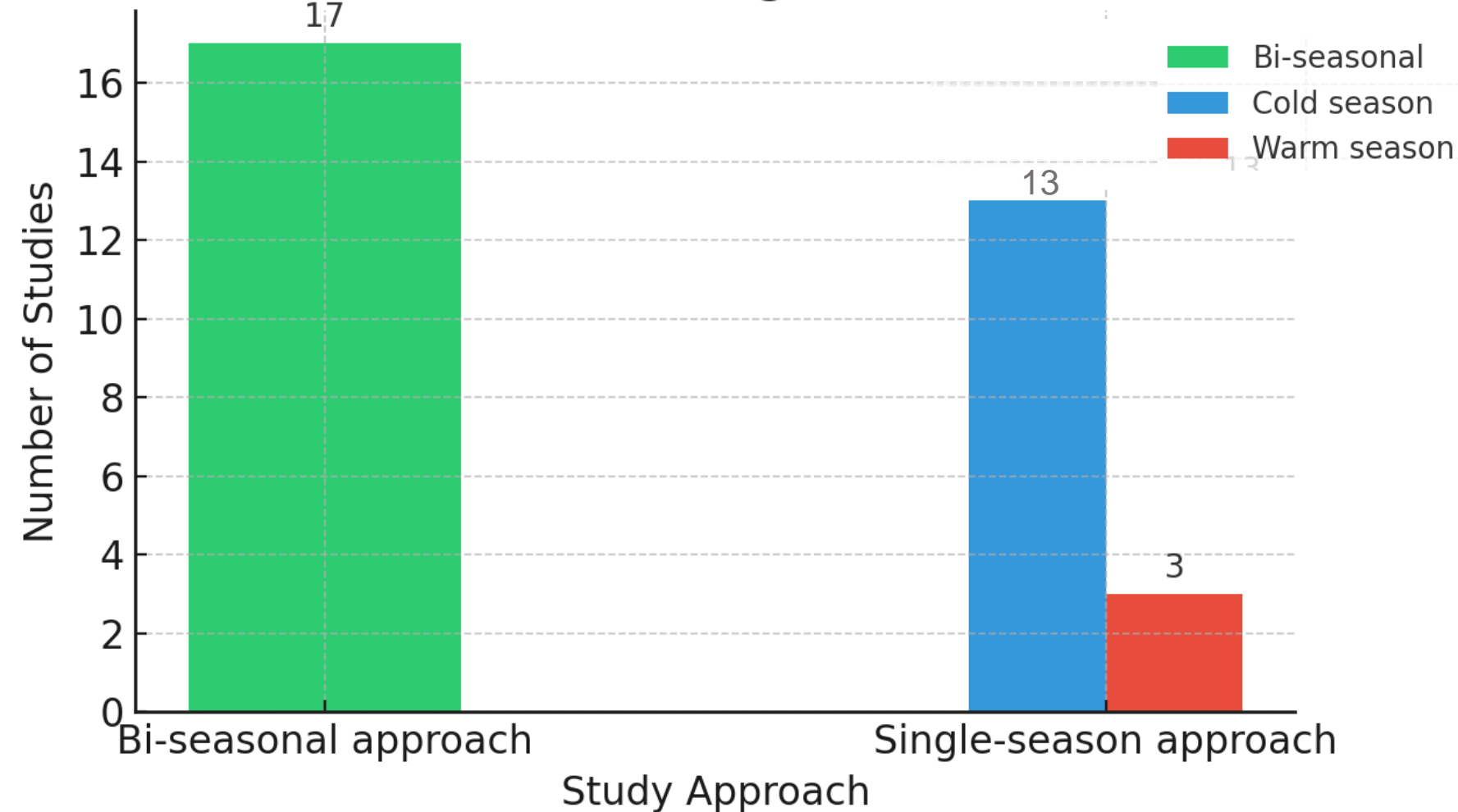
## Oceania

Australia (N=1)



# Seasonal Variations

## Seasonal Focus in Fungal Studies (N=33)



Note: Data from a subset of 33/47 studies that considered seasonal variations.

# Key Factors Affecting Indoor Fungal Concentrations

Health-linked genera fluctuate with PM/temperature  
(Pyrrri et al., 2020)

**Dominant Genera**

10%

Higher concentrations due to poor ventilation  
(Madureira et al., 2014)

**Winter Conditions**

Poor ventilation ↑ risk  
(Ramachandran et al., 2005)

**Ventilation**

20%

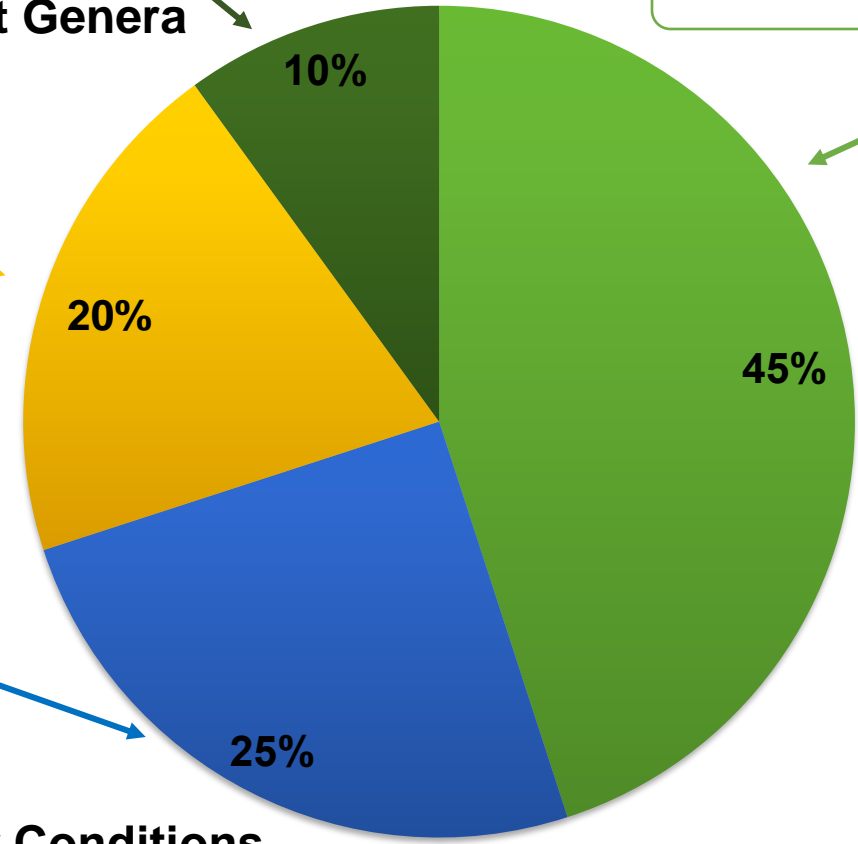
Lower baseline but seasonal genera shifts  
(Minahan et al., 2022)

**Summer Conditions**

25%

45%

**Seasonal variation is a crucial factor in assessing indoor fungal concentrations.**



# Sampling Methods



- 32 out of 47 studies use active sampling methods (with pumps, impactors, filters).
- 15 studies rely solely on passive methods (settled dust, electrostatic cloths, open dishes).
- Only 7 studies combine active and passive methods – a recommended approach for more comprehensive results.
- Active methods: controlled air collection, allow identification of viable particles via culture.
- Passive methods: simple, no equipment needed, suitable for long-term sampling.
- Passive samples require liquid extraction for analysis.
- Combining methods improves data quality and coverage.
- High methodological variability limits cross-study comparisons.

Method Category	Sampling Method/Samplers	References
Passive	Burkard Indoor Recording Air Sampler	Baxi, 2019; Baxi, 2013; Chen, 2014; Su, 2001; Pyri, 2020
	Single-stage microbiologic air impactor (Merck MAS-100)	Cavaleiro Rufo, 2017; Mentese, 2009
	Single-stage Airideal 3P impactor	Fonseca Gabriel, 2021; Madureira, 2014
	Andersen N6 single-stage impactor	Bartlett, 2004; Ramachandran, 2005; Zhang, 2013; Yamamoto, 2015
Impaction	Six-stage Andersen 10-800 impactor	Meklin, 2002; Meklin, 2003; Putus, 2004
	Mattson-Garvin slit-to-agar impactor	Foarde, 2004
	Andersen two-stage cascade impactor	Awad, 2018; Straus, 2003
Active	IOM Inhalable Dust Sampler (SKC Inc.)	Holst, 2016; Würtz, 2005
	Air-O-Cell Cassette (SKC/Zefon)	Santilli, 2003; Godwin, 2007
	Nucleopore filters (0.4 µm)	Kim, 2007; Simoni, 2011
	Millipore cassettes (0.45 µm)	Shin, 2015
	Fine Particle Sampler (PM2.5 filters)	Foarde, 2004
Filter	MCE filter cassettes (0.8 µm)	Ramachandran, 2005; Nieto-Caballero, 2022
	Micro-vacuum sampler (IAQ-1294)	Ramachandran, 2005; Würtz, 2005
	Siemens Super XS vacuum cleaner	Simoni, 2011; Celtik, 2011
	LiT Hummer backpack vacuum	Park, 2022
Vacuum	HVS-3 vacuum sampler	Würtz, 2005
	Generic vacuum cleaners	Chatzidiakou, 2014; Hanson, 2016
Tracer Gas	Tracer-gas decay method (acetone)	Kim, 2007
Other	Sampling pumps (unspecified)	Lee, 2021
	Spot Sampler (Aerosol Devices)	Nieto-Caballero, 2022
Passive	Electrostatic Dust Collectors (EDCs)	Jacobs, 2014; Holst, 2016; Viegas, 2020; Huttunen, 2019; Vomanen-Winqvist, 2020
	Settled dust boxes (SDBs)	Holst, 2016; Huttunen, 2019; Zhang, 2011; Würtz, 2005 (aluminium foil)
	Passive sedimentation (open-dish/gravity plates)	Anduaem, 2019; Aydogdu, 2005; Al-Qurashi, 2007; Hyvonen, 2020; Sauliene, 2023
Swab/Surface Sampling	Surface swabs	Savilahti, 2001; Ejdys, 2011; Hyvonen, 2020; Sauliene, 2023

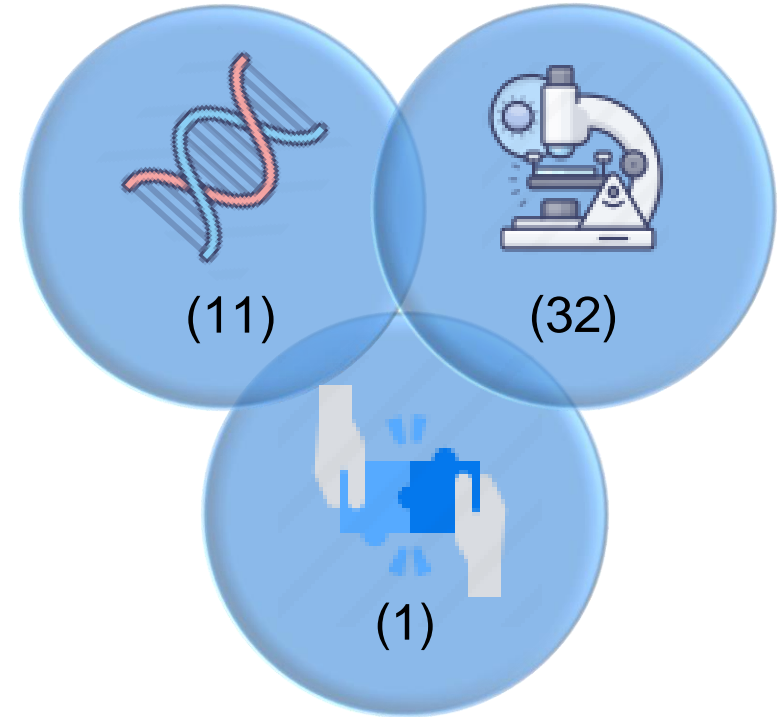
# Analytical Methods

## Culture-based

- ✓ **Morphological visualization** (direct observation of colonies/fungi)
- ✗ **Underestimates non-culturable fungi** (limited to viable samples)

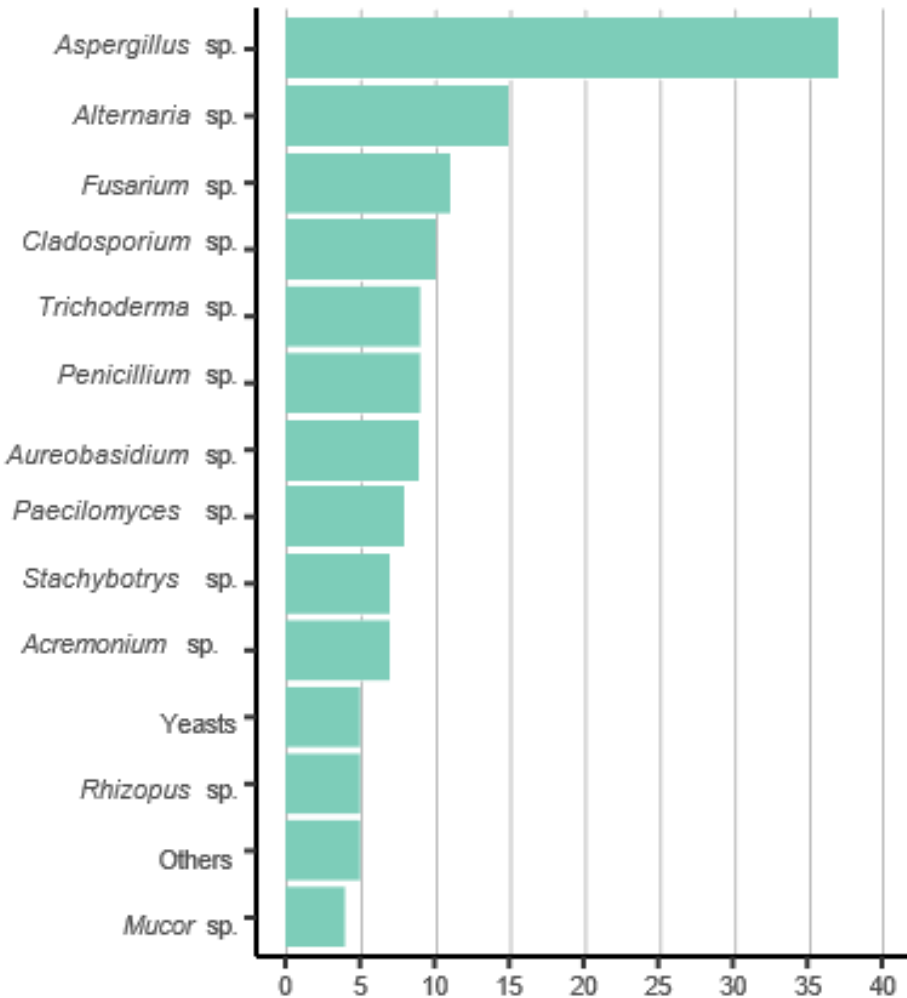
## Molecular

- ✓ **High sensitivity** (detects genetic material even at low levels)
- ✗ **Cannot assess viability** (does not distinguish live vs. dead organisms)



- ✓ **Culture-based methods were most common (68%, 32/47 studies) but underestimate non-culturable fungi.**
- ✓ **Molecular methods (qPCR/DNA sequencing) offered high sensitivity (23%, 11/47) but cannot assess viability and are costly.**
- ✓ **Only 1 study combined culture + molecular methods, revealing a critical gap in hybrid approaches.**
- ✓ **Most studies (64%) relied on single-method analyses, limiting comprehensive risk assessment.**

# Prevalent Fungal Species



## 🌀 *Aspergillus* Dominance

*Aspergillus* was the most prevalent genus, detected in 42/47 studies (89%).

Yet, only 5 studies identified the critical-priority species *A. fumigatus* (WHO, 2022).

## ⚠️ High-Priority Pathogens Underreported

*Fusarium* (11 studies) and *Mucorales* (9 studies)—both WHO high-priority fungi—were far less documented than *Aspergillus*.

## 🏠 Indoor School Environments Harbor Key Genera

11 genera (e.g., *Cladosporium*, *Penicillium*, *Stachybotrys*) were recurrent in elementary schools—potential hotspots for exposure.

## 🔍 Identification Bias Risk

Macroscopically recognizable genera (e.g., *Aspergillus*, *Penicillium*) may skew data, leaving other pathogens understudied.

## 💡 Critical Gap

Low species-level resolution (only 16/42 *Aspergillus* studies specified sections) limits risk assessment accuracy.

# Prevalent Fungal Species

## Most prevalent species

*Aspergillus niger*, *A. fumigatus*, and *A. versicolor* were most frequently detected.

## Identification challenges

Traditional morphology-based classification is insufficient (many species are "cryptic").

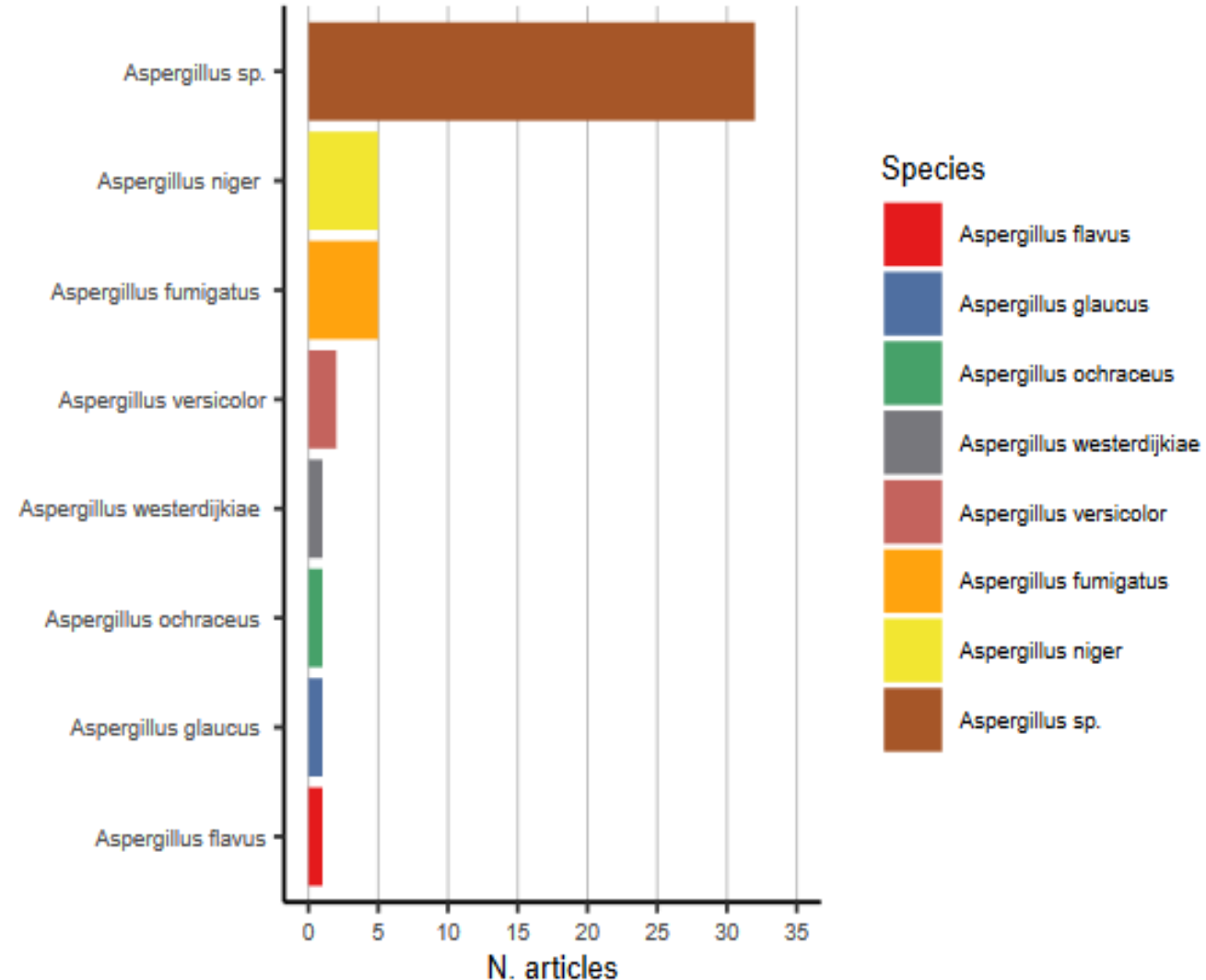
Requires combined methods (morphological + biochemical + genetic) for accurate identification.

## Clinical and public health importance

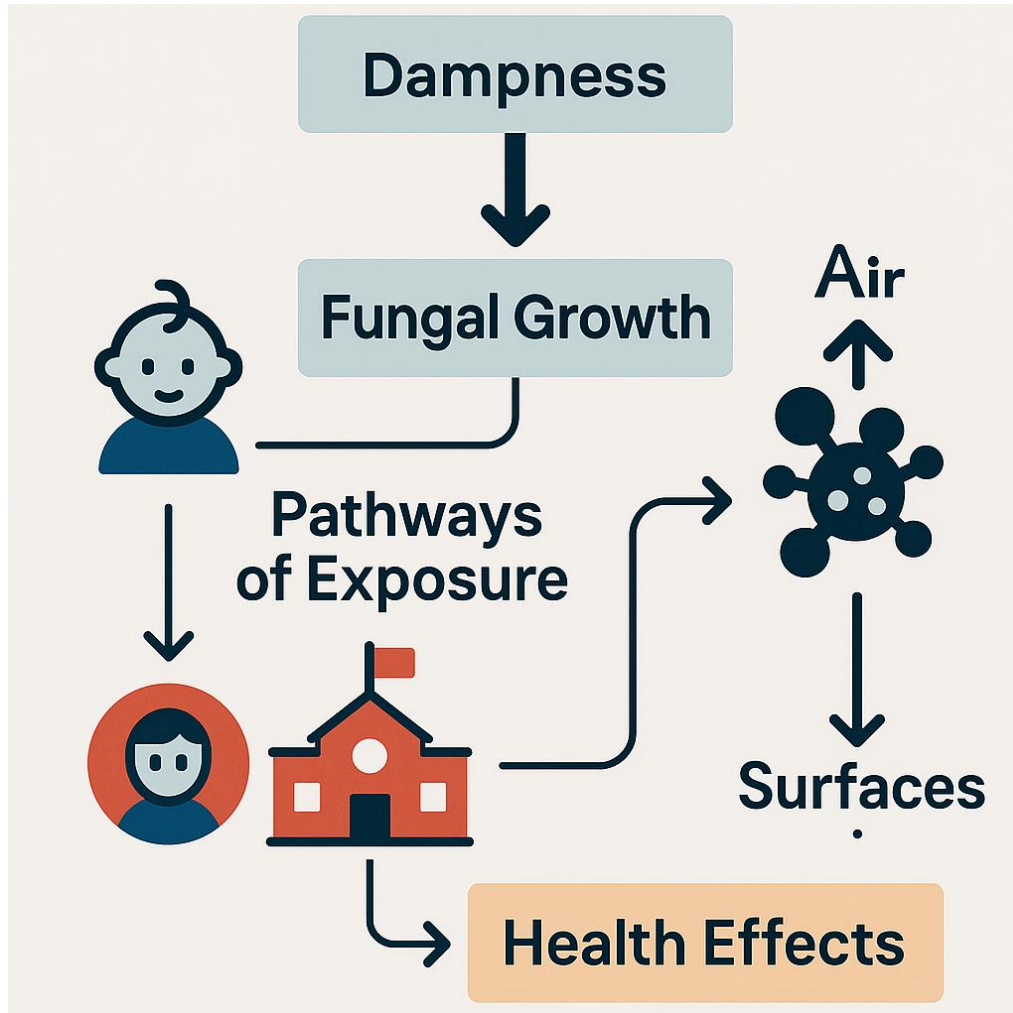
Section-level identification (e.g., *Fumigati*, *Flavi*) is critical for:

Treatment decisions (antifungal resistance)

Infection control and public health interventions



# Health and Structural Impacts



- **Health Risks:** Exposure to fungi (*Aspergillus*, *Penicillium*, *Cladosporium*) in schools is linked to:
  - Respiratory issues (asthma, allergies, infections).
  - Cognitive dysfunction and reduced academic performance.
  - Chronic conditions from mycotoxin exposure (e.g., *Stachybotrys atra*).
- **Structural Damage:** Fungal growth compromises building integrity, especially in damp/moisture-prone areas.
- **Vulnerable Groups:** Children, immunocompromised individuals, and those with pre-existing conditions are at higher risk.

# Limitations



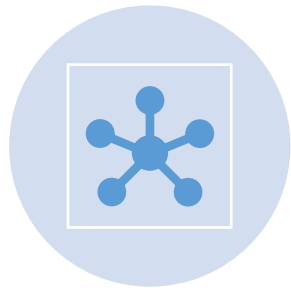
## Methodological Gaps:

- 64% of studies used single-method approaches (culture or molecular), limiting comparability.
- Lack of standardized reporting (e.g., units like CFU/m<sup>3</sup>).

Geographic Bias: 54% of data from Europe; underrepresentation of tropical climates.

Health Data Gaps: Few studies directly link fungal exposure to dose-specific health outcomes.

# Conclusions



## **Standardize Protocols:**

Harmonize sampling (active + passive) and analysis (culture + molecular) methods.



## **Target High-Risk Fungi:**

Prioritize *A. fumigatus*, *Fusarium*, Mucorales, and toxigenic species (*Stachybotrys*, *Aspergillus* section *Flavi*).



## **Seasonal/Geographic Monitoring:**

Address variations in fungal loads (higher in winter due to poor ventilation).



## **Policy Action:**

Integrate fungal monitoring into school health programs and building codes.



# THANK YOU

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